Coating of microscope slides

Gelatine coating (from Amann, 1995):

1) Clean the microscope slides in a solution of ethanolic KOH (10 % KOH in 95 % EtOH) for ~ 1 hour. Air dry.

2) Dip microscope slides into a gelatine solution (0,075 % gelatine, 0,01 % chromium potassium sulphate dodecahydrate (for conservation) at 70º C. Air dry in a vertical position.

3) The coated microscope slides can be stored for longer periods (months) at room temperature.

Poly-L-Lysin coating (e.g. based on SIGMA, product number P8920)

1) Clean microscope slides in acid alcohol (1 % HCl in 70 % EtOH). Air dry over night at room temperature, or at 60º C for 1-2 h. For cleaner microscope slides, flush with air in order to remove drops.

2) Allow diluted poly-L-Lysin (0.01 %, diluted in sterile deionised water) to come to room temperature (100 ml) in coplin jars).

3) Place microscope slides in the coplin jar for 5 min.

4) Drain microscope slides and dry for about 1 h at 60º C or over night at room temperature.

Comment:
- Poly-L-Lysin can be obtained from e.g. Sigma as a 0.1 % (w/v) solution.
- Stock solution may be stored at room temperature until expiration date.
- Diluted solutions should be stored at 4º C and used within 3 months.
- Turbid solutions should of course be discarded.
- The maximum number of microscope slides that can be coated is 900/L of diluted solution.
- The coated microscope slides can be stored for longer periods (months) at room temperature.

Amann, R. 1995. In situ identification of microorganisms by whole cell hybridization with rRNA targeted nucleic acid probes. Methods Molecular Ecology Manual. 3.3.6, 1.